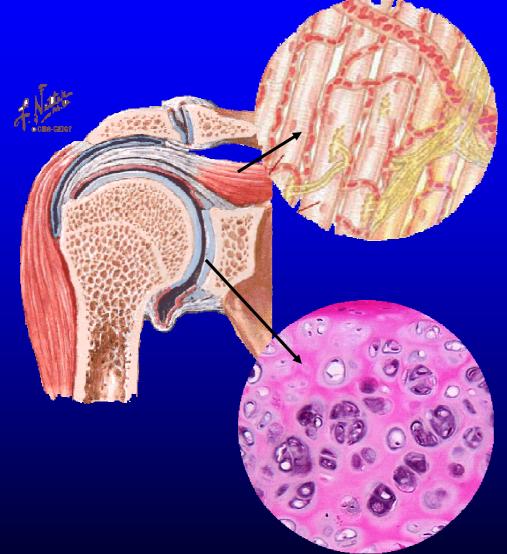


Methods of Biomaterials Testing Lesson 2

Cell Culture and Tissue Engineering



Tissue Consists of Cells



Cells show main reactivity → Use isolated cells for biomaterials testing

Tissue consists of

Cells

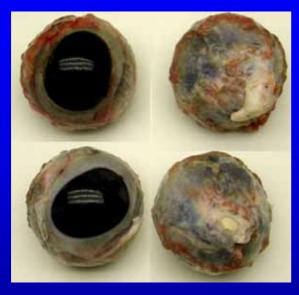
- Specific function
- \rightarrow Parenchym
- Structure
- → Stroma, mesenchym fibroblast/-cyte: tendon, connective tissue osteoblast/-cyte: bone chondroblast/-cyte: cartilage adipocyte: fat tissue

Extracellular matrix

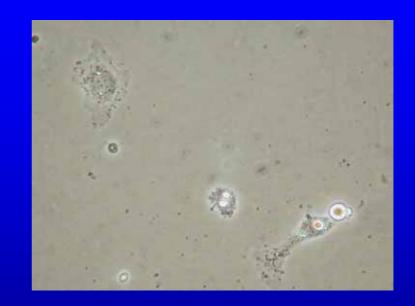
- water
- Collagenes
- Proteogycanes
- Calcium Phosphates
- Soluable substances, mediators

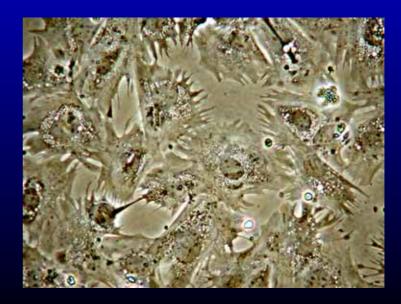


Bovine Retinal Pericytes







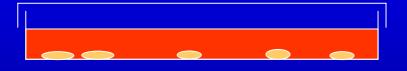




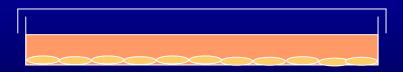
Cell Culture



10⁵ to 10⁶ cells/ml in suspension







Confluent cell layer



Cell Culture Equipment



Cell Culture Medium

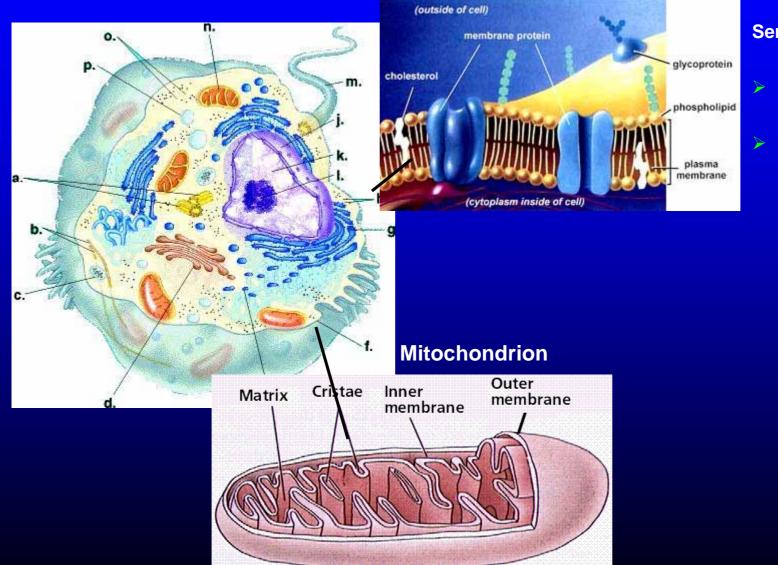
- Nutrients/ Glucose
- Vitamines
- Amino acids
- Salts
 - Osmotic pressure 300mOsm/L
 - pH buffering salts $H_2O + CO_2 \iff H_2CO_3^- + H^+$

Phenol Red

- pH indicator
- **Fetal Bovine Serum**
- Var. Growth factors
- Var. Proteins
- (antibiotics)

Organelles of the Cell

Cell Membrane

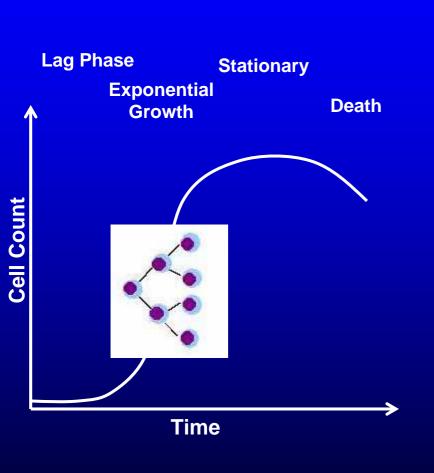


Semipermeable membrane:

- Freely permeable for water
 - Controlled permeability for ions, messengers via gates, pores



Cell Growth



- Lag Phase
 - Low cell growth
 - Cells adapt to the environment
- Exponential growth
 - Maximum cell growth
 - Ideal phase for experiments
- Stationary Phase
 - No more cell growth due to contact inhibition
- Death
 - Death exceeds cell growth

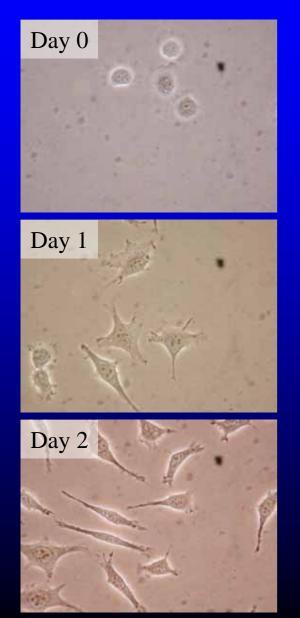


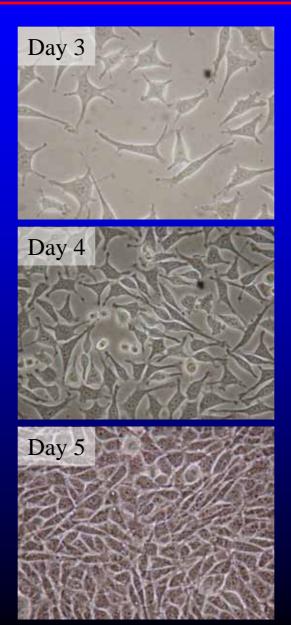
Types of Cell Culture

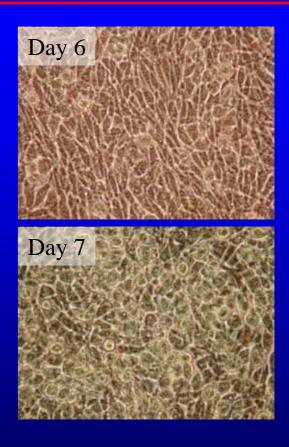
- Culture of Cell Lines
 - commercially available
 - highly standardized
 - immortal
 - as a model more or less far from reality
- Primary Cell Culture
 - Material from individual donors with biological variability
 - Standardized protocols
 - slow growth, limited lifetime
 - closer to reality, interactions between different cells take place
- Tissue Engineering (3 dimensional)
 - little standardization
 - few protocols
 - difficult
 - can be close to reality



Cell Culture – L929

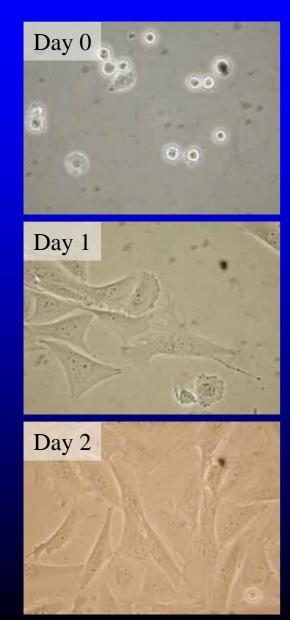


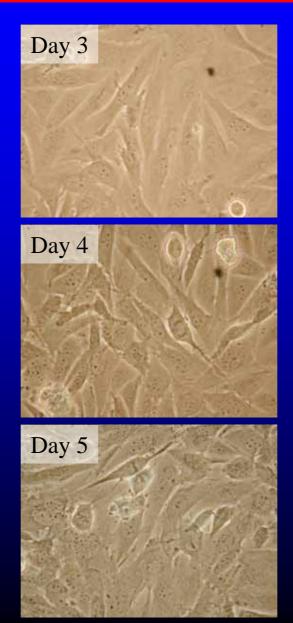


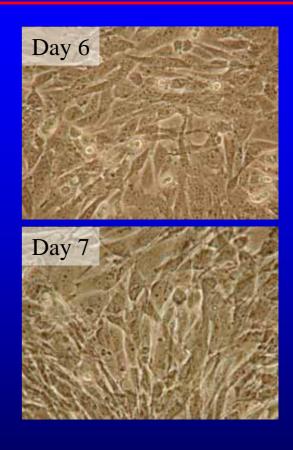




Cell Culture – SAOS-2

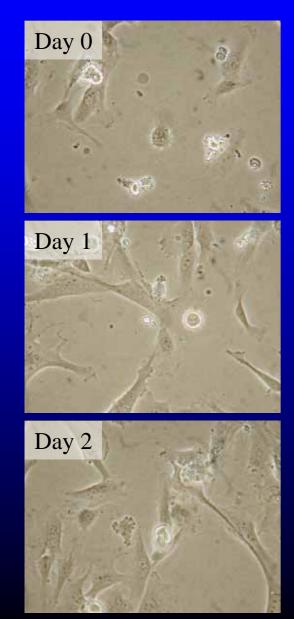


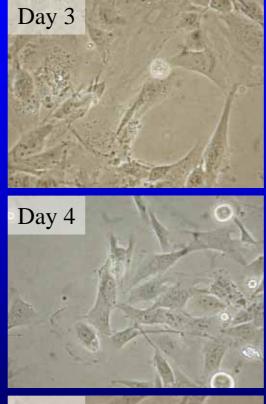


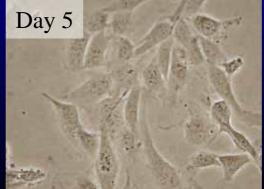


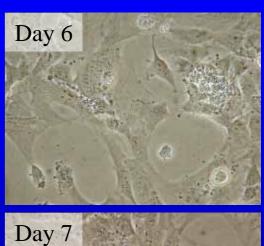


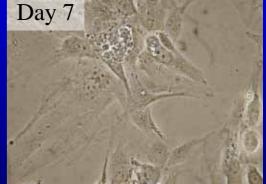
Cell Culture – HMEC-1













Sterile Working

Purpose

- Cell culture has no immune system or other type of immune protection
- Bacterial doubling time is much faster 20min vs. 1 day
- Bacteria/fungus/yeast directly influence cells
- Bacteria/fungus/yeast release toxic substances

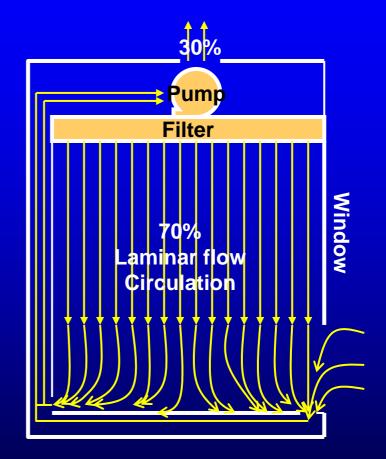
Method

- Desinfection of the Working place
 - 70% Alcohol
- Work on a Laminar Flow Bench
 - Sterile filtered air
- High discipline
- Sterile, single-use equipment
- Sterilize other equipment
 - Dry heat
 - 180°C, 2 hours
 - Autoclavation
 - 121°C, 2bar H₂O vapour
 - Ethylene oxide, Formaldehyde
 - Irradiation
- Sterilize liquids
 - Autoclavation if heat resistant
 - Filter sterilization
- (antibotics)



Laminar Flow Bench (S2)







Autoclave





Sadvantages of standard Cell Culture

- Only one cell type
 - No communication/ interaction of cells
 - Not the original composition of the extracellular matrix
 - →Co-culture
 - →Animal experiment
- Only two-dimensional cell growth
 - \rightarrow Tissue Engineering: seeding cells on a 3D scaffold
- Static conditions
 - No load, no shear forces
 - Dynamic cell cultures, culture in flow chambers...



Cell Co-Culture I

Grow several cell types together in one medium <u>Purpose</u>: Check the influence of one cell-types reaction on an other cell type or use the environment of one cell type, which is necessary for the other

Problems:

- Because of the different speed of growth of different cells, one soon will overgrow the other
- Cells cannot be identified any more

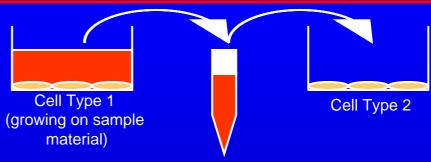


Cell Co-Culture II

- Transfer of supernatant
 - Convenient
 - Reliable
 - One-way communication only

Co-Culture with membranes, insets

- Direct contact possible
- Bilateral communication
- Expensive
- Low number of cells
- Problems with oxygen supply
- Compared to natural conditions still not the original Extracellular matrix
- Co-culture with Gamma-irradiated cells
 - Mainly for nursing/ feeder cells, which provide environment for other cells

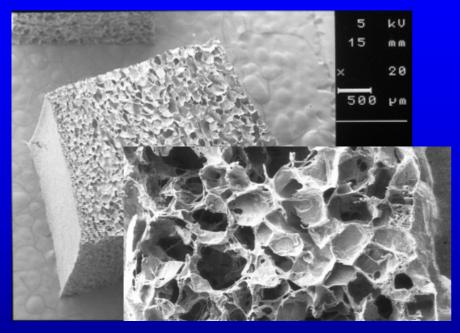


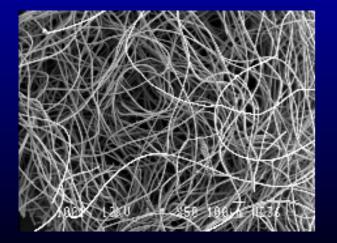
Filter or centrifuge





Tissue Engineering





Principle

 Seed cells on a porous 3D scaffold

Technical Problems

- Limited penetration of the cells
- Limited perfusion of the scaffold
 - Oxygen
 - Nutrients
- Perfusion becomes worse with cell growth

Ways to overcome Problems

- Perfusion chambers
- Seeding endothelial cells first